

PLATYHELMINTHES FROM SALT MARSHES OF COOMERA RIVER, SOUTHEASTERN QUEENSLAND, AUSTRALIA

ANNO FAUBEL AND BRONWYN CAMERON

Faubel, A. & Cameron, B. 2001 06 30: Turbellaria from salt marshes of Coomera River, southeastern Queensland, Australia. *Memoirs of the Queensland Museum* 46(2): 511-519. Brisbane. ISSN 0079-8835.

Three new species, *Childiana coomerensis* gen. et sp. nov. (Acoela), *Macrostomum greenwoodi* sp. nov. and *Macrostomum coomerensis* sp. nov. (Macrostomida) are described from the brackish water estuarine area of the Coomera river, Gold Coast, Queensland, Australia. □ *Taxonomy, morphology, flatworms, Acoela, Macrostomida.*

Dr A. Faubel, Institut für Hydrobiologie und Fischereiwissenschaft, Universität Hamburg, Zeiseweg 9, D-22765 Hamburg, Germany (e-mail: faubel@uni-hamburg.de); B. Cameron, The University of Queensland, St Lucia 4072, Australia: 14 March 2000.

The Australian fauna of Acoela and Macrostomida of limnetic, brackish and marine habitats is not very well known. Over a long time the only study known on Acoela was carried out by Haswell (1905) on *Heterochaerus australis*. More recently, however, more intensive studies have been started by Trench & Winsor 1987 on *Amphiscolops* sp. and *Haplodiscus* sp. (in part *Waminoa litus* Winsor, 1990), Winsor 1988 on *Wulguru cuspidata*, Winsor 1990 on *Convolutriloba hastifera*, *Waminoa litus*, *W.* sp. 1, and *W.* sp. 2.

A first census of macrostomid species of Australia by Faubel et al. (1994) listed 6 species. Of these *Macrostomum tuba* v. Graff, 1882 and *Promacrostomum palum* Sluys, 1986 are known from freshwater environments. *Dunwichia arenosa* Faubel, Blome & Cannon, 1994, *Bradburia australiensis* Faubel, Blome & Cannon, 1994, *Macrostomum australiense* Faubel, Blome & Cannon, 1994, and *Macrostomum* sp. Faubel, Blome & Cannon, 1994, however, are all known from eulittoral habitats of sandy beaches of Stradbroke Island, South Queensland.

In the present paper one acocloous species and two macrostomid species from brackish water habitats are described.

METHODS

For studies on meiobenthos, sediment was qualitatively collected from the Coomera salt-marsh pools. The freshly collected sediment was transferred into a glass tank in a constant temperature 1.00m approximating ambient conditions (Salinity ca 30 ppt, 27°C). Habitat water was added to the sediment tank, to a depth of about 15cm. A series of 60 watt light bulbs

were suspended above the tank to attract organisms into the water column. Every hour, the tank was siphoned through a 63µm sieve to collect emerged fauna. The defaunated water was then returned to the tank and the extraction process repeated until the majority of the organisms had been collected. Sexually mature specimens of Turbellaria were studied alive and in squash preparation, i.e. flattened under the increasing pressure of the coverslip as the preparation dried. Measurements of living organisms (m.l.s.) were made from squashed ones. These measurements are given in parentheses in the running text. All other measurements were made from sections of the holotype. For histological observation specimens were relaxed in 7% MgCl₂ and fixed in Bouin's fixative. Specimens were embedded in Paraplast plus (Reichert & Junk) and cut sagittally at 3.0µm and stained with haematoxylin-eosin according to Mayer. Types are deposited in the Queensland Museum, Brisbane.

SITE DESCRIPTION

Coomera salt-marsh (27°54'S, 153°17'E) was chosen as a site for turbellarian collection. For the past 4 years this site had been excluded from the broad-scale mosquito control measures that typically occur in southeast Queensland. It was therefore considered to be a relatively pristine site.

Coomera salt-marsh is only inundated by tides of 2.4m or more above datum. Salinity ranges from 25-38 ppt. The salt-marsh pools are 2-7m² in area and are bordered by dense clumps of *Sporobolus virginicus* (Kunth.) the dominant marsh-grass, and *Sarcocornia quinqueflora* Bunge (ex. Ung. Stern) a sprawling herbaceous

halophyte. The sediment consists of much highly organic mud with negligible amounts of sand.

SYSTEMATICS

Order ACOELA Uljanin, 1870
Family CHILDIIDAE Dörjes, 1968

Childianea gen. nov.

DIAGNOSIS. Childiidae with frontal organ. Body wall musculature with outer circular and inner longitudinal muscle fibers. Separate genital openings. Seminal bursa with vagina and cellular bursa mouth directed anteriorly. Paired ovaries and testes. Male copulatory apparatus without seminal vesicle; proximal ejaculatory duct with sclerotised layer working as penial stylet; distal ejaculatory duct with prostatic glands. False seminal vesicles present.

TYPE SPECIES. *Childianea coomerensis* sp. nov.

ETYMOLOGY. Derived from the generic name *Childia*.

KEY TO THE GENERA OF THE CHILDIIDAE

1. with single male copulatory organ 3
with several male copulatory organs 2
2. with 2 male copulatory organs . . . *Childia* v. Graff, 1910
with 4 male copulatory organs
. *Tetraposthia* An Der Lan, 1936
- 3(1). . . with single male copulatory organ; without seminal
bursa 4
with seminal bursa 6
4. without male atrium or very short 5
with long male atrium *Atriofronta* Dörjes, 1968
5. copulatory organ with sclerotised spines
. *Actinoposthia* An Der Lan, 1936
copulatory organ with rosette like muscular elements
encasing the seminal vesicle
. *Paraproporus* Westblad, 1945
- 6(3). with seminal bursa; body muscle wall with outer circular
and inner longitudinal muscle fibers. 7
body muscle wall with inverse muscle layers
. *Paraphanostoma* Steinhöck, 1931
7. with bursa nozzle. 8
without bursa nozzle . . . *Pseudactinoposthia* Dörjes, 1968
8. bursa nozzle directed anteriorly or caudad 9
bursa nozzle invaginated in the seminal bursa
. *Pelophila* Dörjes, 1968
9. bursa nozzle cellular 10
bursa nozzle sclerotised . . . *Philactinoposthia* Dörjes, 1968
10. male complex with prostatic glands, without seminal
vesicle *Childianea* gen. nov.
male complex with seminal vesicle, without prostatic
glands. *Archactinoposthia* Dörjes, 1968

Childianea coomerensis sp. nov. (Figs 1A-B, 2)

ETYMOLOGY. From the Coomera River estuary on the Gold Coast of Queensland.

MATERIAL. HOLOTYPE. Coomera salt-marsh (27° 4'S, 153° 17'E), Gold Coast, southeast Queensland, QM G217362, 2 specimens, May 1998 leg. B. Cameron.

DESCRIPTION. Length of body of living sexually mature specimens up to 0.9mm, when extended and gliding; maximum width up to 0.18mm in second half of body, area of growing oocytes. Body spindle-shaped with rounded frontal and rear body end, dorso-ventrally flattened. Posterior body end with a small vacuole being horseshoe-like (Fig. 1A). In transmitted light, colour of body yellowish but digestive parenchyme and lumen bright yellow through ingested diatoms. Glandular ducts of frontal organ open to exterior through several pores at the frontal end, the glands of which lie posterior to the statocyst. Secretion of the glands coarsely granulated. Statocyst embedded within the medio-frontal brain, about 55µm (m.l.s.) distant from anterior body end. Epidermal cells 2.0 - 2.5µm high, with intraepithelial nuclei; rhabdoids lacking. Epidermis completely covered with cilia 3.5 - 4.0µm in length. Body wall musculature with faint outer circular and inner longitudinal fibers. Dorsal and ventral subepidermal hyaline glands open to the exterior throughout body surface, more abundant lateral. Digestive lumen bordered by nucleus-rich parenchymal tissue which starts immediately at level of frontal glands and fills the whole mid-body covering the ovaries dorsal. Gut lumen filled with diatoms and detritus. Mouth ventral behind mid-body, about 400µm (m.l.s.) distant from frontal body end.

Reproductive system (Fig. 2). Lateral testes situated about 180µm (m.l.s.) from anterior body end. Spermatids penetrate the parenchyma on both sides lateral of the ovaries. Anterior to the transverse level of the male complex the spermatids accumulate in two false seminal vesicles which enter the common proximal ejaculatory duct in common. True seminal vesicle lacking. Ejaculatory duct is closed by a sphincter at junction of the vasa deferentia. The ventral male pore (Fig. 1B,2), 32µm distant from rear body end, leads to the bulbous pyriform copulatory organ. Muscle bulb a highly muscular complex 43µm in length with central ejaculatory duct (Fig. 2). The muscle fibers run more or less

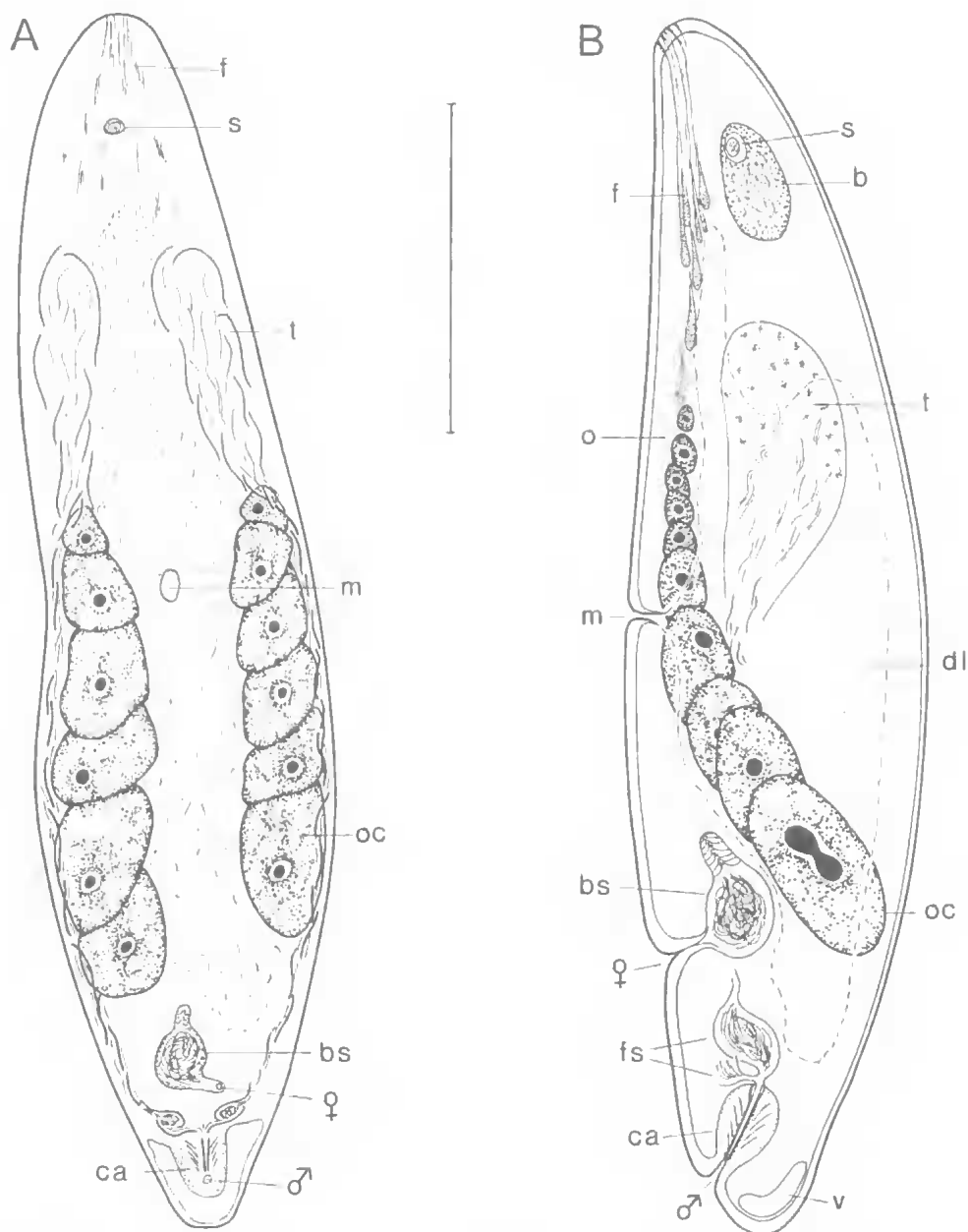


FIG. 1. *Childianea coomerensis*, gen. et sp. nov. A, dorsal view in squash preparation. B, diagrammatic sagittal reconstruction after serial sections. Scale: 200 μ m. (b = brain, bs = seminal bursa, bm = mouth of bursa, ca = copulatory apparatus, dl = digestive lumen, f = frontal organ, fs = false seminal vesicle, m = mouth, o = ovary, oc = oocyte, s = statocyst, t = testis, v = vacuole, ♂ = male gonopore, ♀ = female gonopore).

parallel, inclining toward the ejaculatory duct, and obviously function as retractors during copulation. Ejaculatory duct tripartite; proximal part of duct (Fig. 2) about 10 μ m long, highly sclerotised forming a seal controlling sperm ejaculation; central one, 31 μ m long, lined by a

sclerotised epithelial layer looking stylet-like; distal part is 12 μ m long, not ciliated, and of prostatic function. Cell bodies of epithelial layer very elongated, enclose copulatory complex (Fig. 2).

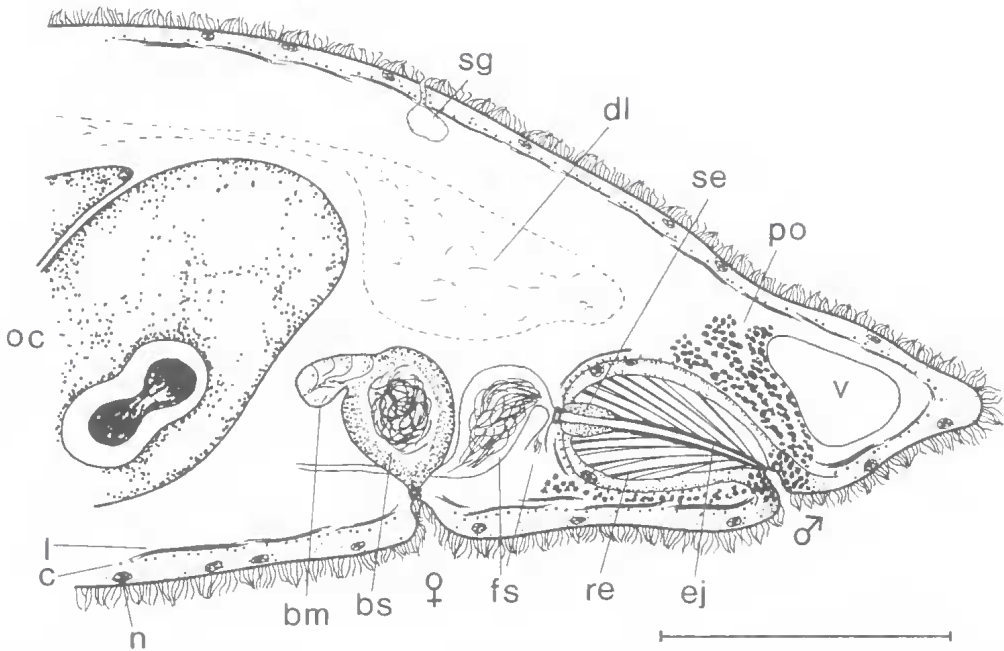


FIG. 2. *Childianea coomerensis*, gen. et sp. nov.; sagittal reconstruction of the male and female complex. Scale: 50 μ m. (bs = seminal bursa, bm = mouth of bursa, c = circular muscles, dl = digestive lumen, ej = ejaculatory duct, fs = false seminal vesicle, l = longitudinal muscles, n = nucleus, oc = oocyte, po = prostatoid glands, re = retractor muscles, se = seal, sg = subepidermal gland, v = vacuole, ♂ = male gonopore, ♀ = female gonopore).

Female system with paired ovaries and a seminal bursa. Oogonia of each ovary lie ventro-lateral, 190-200 μ m behind statocyst (Fig. 1B). A line of growing oocytes runs latero-caudad. Hindmost oocytes are about 170 μ m (m.l.s.) distant from posterior body end. Seminal bursa with a distal ciliated atrium (7.5 μ m long), a central vesicle filled with sperm (20 μ m in diameter), and a cellular mouth piece 13 μ m long. The atrium rises dorsad, 65 μ m anterior to male pore, widening proximad to form a vesicle filled with sperm. The cellular bursa mouth piece is directed anteriad, reaching central the hindmost part of the oocytes. Aperture between atrium and vesicle of bursa is closed by a sphincter.

DIAGNOSIS. With characters of the genus.

DISCUSSION. The family Childiidae was established and discussed in detail by Dörjes 1968. Up to now the family contains 10 genera. Dörjes included in this family all these genera of Acoela which have species with a sclerotised (cuticular according to Dörjes 1968), muscular, or cellular penis plug which is never invaginated in the seminal vesicle. The new species *Childianea coomerensis* is characterized by a sclerotised ejaculatory duct obviously working

as a pointed stylet. On the basis of this character the species belongs to the Childiidae. *Coomera coomerensis*, however, differs from all species of the known genera of the family Childiidae in the presence of prostatic glands entering the distal ejaculatory duct. In combination with the characters of presence of a seminal bursa with a cellular spermiducal duct directed frontad and absence of a true seminal vesicle, establishment of monotypic *Childianea* and its species *C. coomerensis* sp. nov. is justified within the family Childiidae.

Order MACROSTOMIDA, Meixner, 1924
Family MACROSTOMIDAE Van Beneden,
1870

Macrostomum greenwoodi sp. nov.
(Fig. 3)

ETYMOLOGY. Named in honour of Prof. J. Greenwood, University of Queensland.

MATERIAL. HOLOTYPE. Coomera salt-marsh (27°54'S, 153°17'E), Gold Coast, southeast Queensland, QMG217363, 2 specimens, April 1998 leg. B. Cameron.

DESCRIPTION. Length of body of living sexually mature specimens up to 1.3mm, when

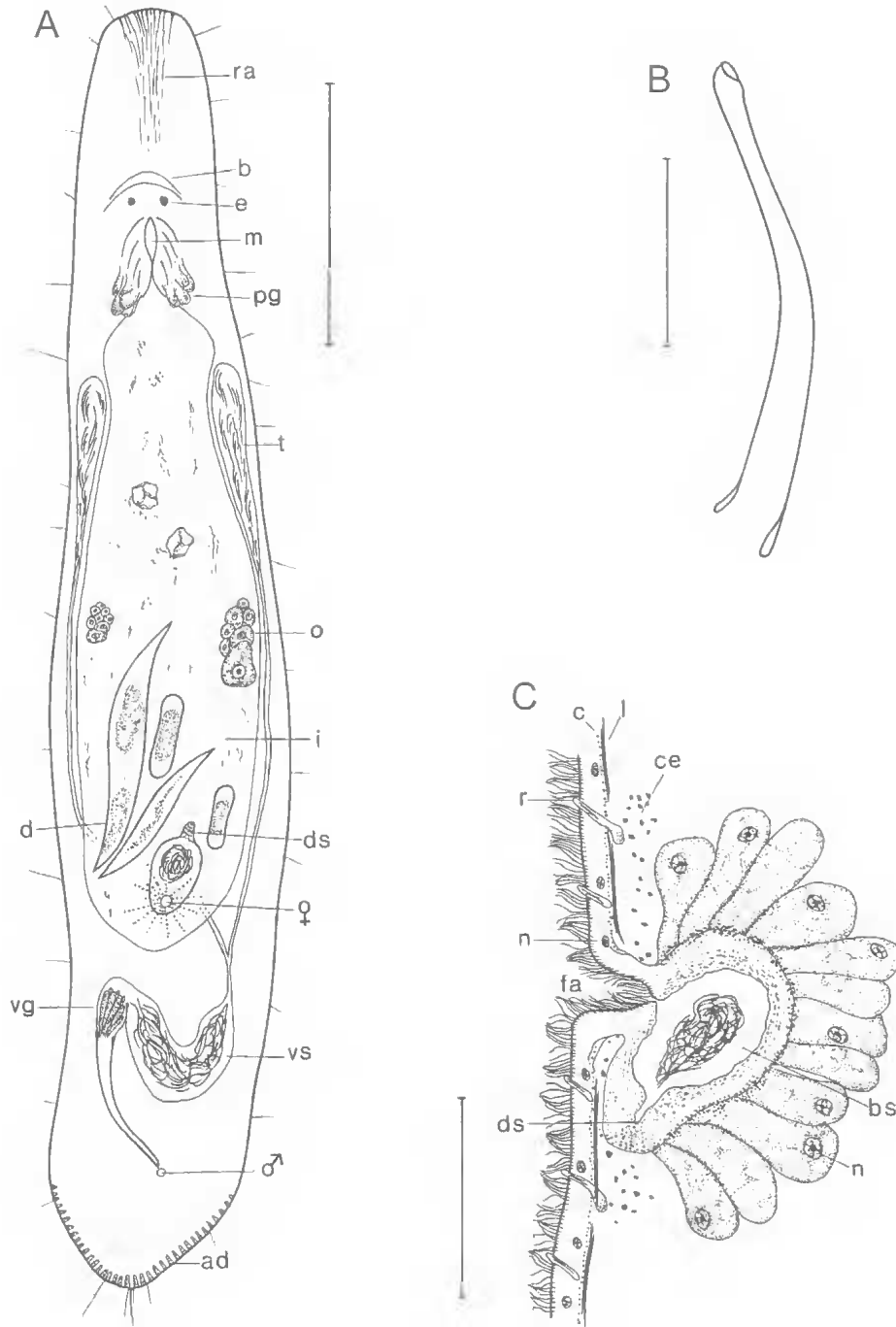


FIG. 3. *Macrostomum greenwoodi*, sp. nov. A, dorsal view in squash preparation. B, male stylet. C, sagittal reconstruction of the female organ. Scales: A, 200µm; B, 20µm; C, 50µm. (ad = adhesive glands, b = brain, bs = seminal bursa, c = circular muscles, ce = cement glands, d = diatoms, ds = spermiducal duct, e = eyes, fa = female atrium, i = intestine, l = longitudinal muscles, m = mouth, n = nucleus, o = ovary, pg = pharyngeal glands, r = rhabdite, ra = rhammites, t = testis, vg = vesicula granulorum, vs = seminal vesicle, ♂ = male gonopore, ♀ = female gonopore).

extended and gliding; maximum width up to 0.18mm in mid-body. Outline of body with characteristic macrostomid like anterior end; posterior end rounded, provided with strong adhesive glands (Fig. 3A). Around margin weak sensory hairs of varying length and stiffness present. In incident light body brownish with darker contrasting oocytes in the posterior part of body; gut intensively coloured yellow based on the uptake and consumption of bright yellow diatoms. Eyes present, 175 μ m (m.l.s.) far from anterior margin of the body. Crescentic brain 169 μ m (m.l.s.) distant from anterior end. Rhammite glands open ventro-frontal through the anterior margin of body, their cells extending posteriad up to the level of the pharynx simplex. In mid-body cells of epidermis 4.6 μ m high covered with 4.0 to 4.6 μ m (ventral) and 3.4 to 3.9 μ m (dorsal) long cilia. Intraepithelial nuclei present; rhabdites about 9.0 μ m long, the cell bodies of which subepidermal, up to 11.3 μ m long. Rhabdites distributed in longitudinal rows over the dorsal and ventral body surface. Body muscle wall with outer circular and inner longitudinal muscle fibers. Basal membrane not observed. Digestive system with pharynx simplex pierced by extrapharyngeal glands and large intestine but less numerous ciliated than pharynx. The intestine fills the median parts over the ovaries, oocytes, and seminal bursa and extends caudad up to the seminal vesicle. Mouth behind brain, 210 μ m (m.l.s.) distant from anterior end. The species feeds on diatoms and detritus particles. Ingested sand grains covered with detritus are abundant. Excretory system not observed, probably absent. Male and female gonopores separate, 185 μ m (m.l.s.) distant from each other.

Reproductive system (Fig. 3). The male system is typically macrostomid like. It consists of bilateral testes, vasa deferentia running caudad to either side of the intestine and ovaries, a common vas deferens, a seminal vesicle, a prostatic vesicle, and a male stylet which projects into the short male antrum. The stylet depicted in Fig. 3A, is 98.3 μ m long; the proximal opening is 12.0 μ m in diameter; the distal opening is subterminal. The male pore is 105 μ m (m.l.s.) distant from rear body end. The antrum rising dorsad is 10.8 μ m long.

The female system consists of bilateral ovaries and a seminal bursa. The ovaries forming lateral fields of oogonia, lie behind the germinative zone of the testes in mid-body (Fig. 3A), generating caudad large oocytes. The seminal bursa consists

of a ciliated atrium (11.4 μ m long), a bursal vesicle (37.9 μ m long), and a spermatid duct (12.8 μ m long). The bursal vesicle is totally lined with elongated cells. The proper epithelium of the bursa looks syncytially in which the surrounding cells (up to 35 μ m) excrete obviously coarsely granulated secretion. Cement glands surrounding the female pore, discharge their secretion into the female atrium.

DIAGNOSIS. The species is diagnosed on the outline of the male stylet (Fig. 3B).

***Macrostomum coomerensis* sp. nov.**
(Fig. 4)

ETYMOLOGY. From the Coomera river of the Gold Coast of Queensland.

MATERIAL. HOLOTYPE. Coomera salt-marsh (27°54'S, 153°17'E), Gold Coast, southeast Queensland, QM G217364, 2 PARATYPES. Same data, QM G217365-366, numerous specimens, April 1998 leg. B. Cameron.

DESCRIPTION. Length of body of living sexually mature specimens up to 1.4mm, when extended and gliding; maximum width up to 0.25mm in anterior part of second body-half, area of oocytes (Fig. 4A). Anterior and posterior body end rounded. Marginal hairs or spines absent. Frontal glands present, not reaching anterior level of brain. Eyes present immediately behind crescentic brain, 125 μ m (m.l.s.) distant from anterior margin of body. In incident light colour brownish yellow with contrasting darker lateral ovaries. In transmitted light body greyish translucent with yellowish digestive system. Mouth opening immediately caudal of eyes. From mouth the pharynx simplex rises dorso-caudad and receives openings of digestive glands. Intestine with less numerous cilia. The gut fills the median parts of the body extending caudad over ovaries and oocytes, ending up at the anterior level of the last oocytes. Rhabdites evenly distributed in serial lines over the body. Ventral rhabdites 7.5 μ m and dorsal ones 9.4 μ m long. Rear dorsal body end with dense aggregates of large rhabdites (10.7 μ m long). Ventral epidermis is 5.7 μ m thick and dorsal one is 2.8 μ m thick, entirely covered with 5.7 μ m long cilia; epithelial nuclei subepidermal. Basal membrane not observed. Body muscle wall with outer circular and inner longitudinal muscle fibers. Excretory system not observed, probably absent. Gonopores separate, 65 μ m distant from each other. Bilateral testes anterior to ovaries; last in mid-body.

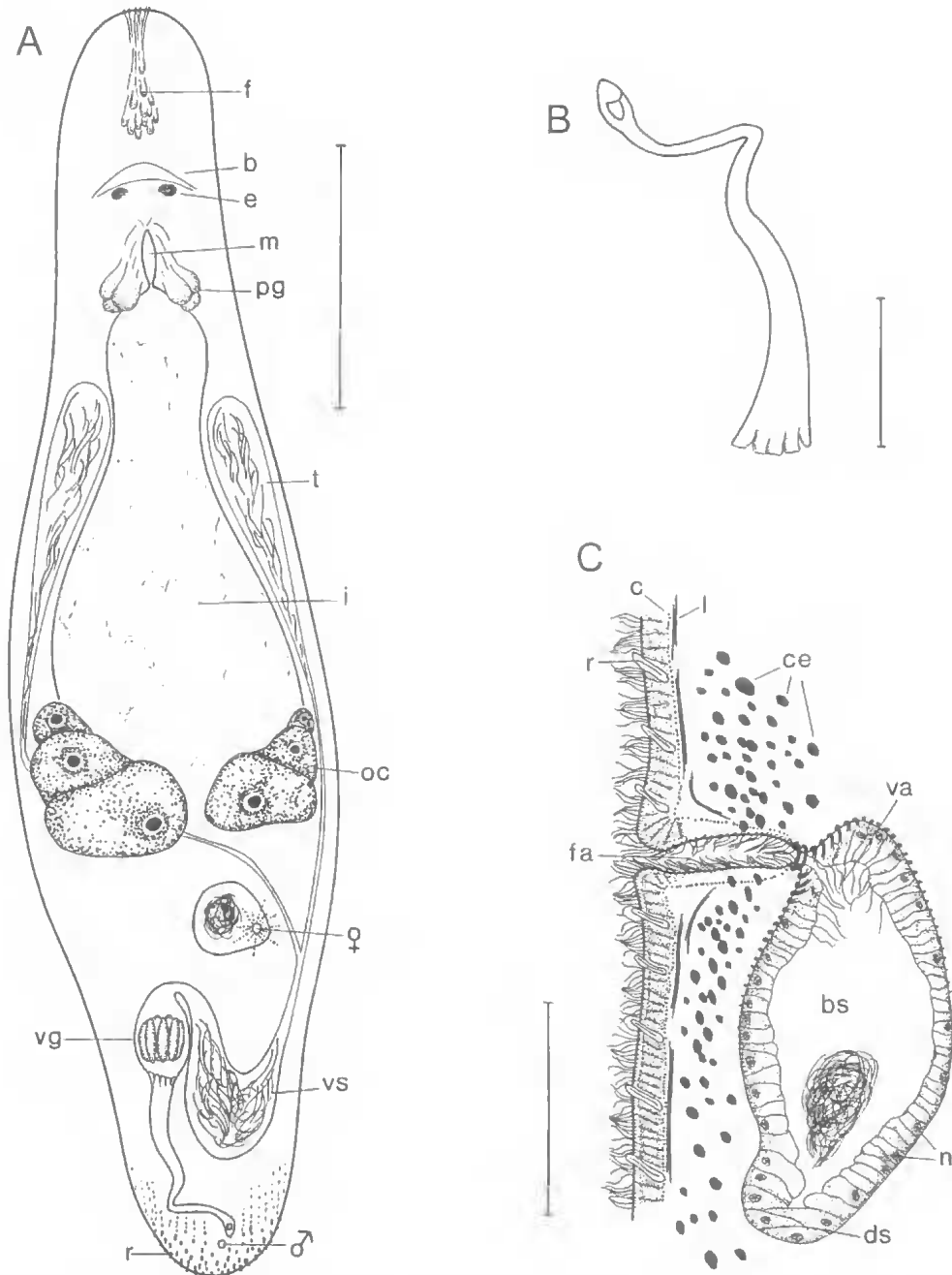


FIG. 4. *Macrostomum coomerensis*, sp. nov.; A, dorsal view in squash preparation. B, male stylet. C, sagittal reconstruction of the female organ. Scales: A, 200µm; B,C, 50µm. (b = brain, bs = seminal bursa, c = circular muscles, ce = cement glands, ds = spermiducal duct, e = eyes, f = frontal organ, fa = female atrium, i = intestine, l = longitudinal muscles, m = mouth, n = nucleus, o = ovary, pg = pharyngeal glands, r = rhabdite, t = testis, va = vagina, vg = vesicula granulorum, vs = seminal vesicle, ♂ = male gonopore, ♀ = female gonopore).

Reproductive system (Fig. 4). The male system is typically macrostomid like. It consists of bilateral testes, vasa deferentia running caudad to either side of the intestine and ovaries, a common vas

deferens, a seminal vesicle, a prostatic vesicle, and a male stylet which projects into the male antrum. The sac-like testes are located 100 to 110 μm (m.l.s.) behind the pharynx. Latero-caudal of the female bursa the vasa deferentia unite to a common vas deferens on the left side before entering the seminal vesicle. The distal part of the seminal vesicle, the prostatic vesicle, and the proximal part of the penial sheath in which the stylet rests, are covered with a complicated system of strong muscle fibers. These fibers are responsible for protrusion and retraction of the stylet and ejaculation of sperm into the female system. The distal part of the prostatic vesicle is encased in the proximal portion of the stylet being slightly funnel-like extended. The stylet is 125 μm (m.l.s.) long; the distal two thirds are spiralled as outlined in Fig. 4B. The distal tip of the stylet is broadend, the opening of which lies subterminally. The male pore, 42.7 μm distant from the rear body end leads antero-dorsad to a rather long male antrum (15.2 μm).

The female system consists of bilateral ovaries and a seminal bursa. The oogonia generate laterally immediately behind the testes and are growing up caudad forming oocytes on each side of the body. The female genital pore is 280 μm (m.l.s.) distant from the caudal body end. The female atrium (43.5 μm long) entered by cement glands, is outlined with cilia and rises dorsad. The transition to the vagina is closed by a sphincter. The vagina, up to 31.5 μm long, ciliated, extends frontad to form a bursal vesicle filled with sperm. The cilia of the vagina up to 13.0 μm long. Anteriad of the bursa a spermiducal duct (24 μm long) is present. The spermiducal duct reaches up mediad to the level of the last oocytes. The epithelia of the vagina, the bursal vesicle, and the spermiducal duct are cellular with intraepithelial nuclei.

DIAGNOSIS. The species is diagnosed on the outline of the male stylet (Fig. 4).

DISCUSSION. Macrostromidae currently contains 11 genera. The dominant taxonomic characters concern the male copulatory apparatus: a distally armed ejaculatory duct (stylet), or an unarmed cirrus. Genera having a penial stylet are *Macrostromum* Schmidt, 1848, *Omalostomum* Van Beneden, 1870, *Promacrostromum* An Der Lan, 1939, *Archimacrostromum* Ferguson, 1954, *Bradynectes* Rieger, 1971, and *Bradburia* Faubel, Blome & Cannon, 1994. Of these genera, *Macrostromum* has the most species (127).

Beklemishev (1951) has assigned the species of *Macrostromum* to 3 taxonomic groups as follows: *Macrostromum hystericum* group (stylets being funnel-like proximally, with tapered hook-like shape distally), *Macrostromum orthostylum* group (stylets being straight or curved, distad evenly tapering), and *Macrostromum tuba* group (stylets having a distal characteristic *Macrostromum tuba*-like enlargement, with the opening of the ejaculatory duct in the centre of the tip, or subterminally). Although these groupings are useful in a taxonomic sense, they should not be regarded as phyletic groupings because there are different lines of the development of the female genital organs within each group. A revision of the taxon Macrostromida is in preparation by the first author. Based on the distal features of the stylet, *Macrostromum coomerensis* belongs to the *Macrostromum tuba* group and *Macrostromum greenwoodi* to the *Macrostromum orthostylum* group.

Macrostromum coomerensis is characterised by having the male stylet formed like a cork-screw with a distal enlargement, and a subterminal opening of the ejaculatory duct. On the basis of these features the species belongs to the *Macrostromum tuba* group. *Macrostromum* species with a spiral stylet are: *M. leptos* An Der Lan, 1939, *M. lewisi* Ferguson, 1939, *M. reynoldsi* Ferguson, 1939, *M. riedeli* Ferguson, 1940, *M. delphax* Marcus, 1946, *M. contortum* Beklemishev, 1951, *M. phytophilum* Beklemishev, 1951, *M. phocorum* Marcus, 1954, *M. spirale* Ax, 1956, *M. poznanienne* Kolasa, 1973, *M. bicurvistyla* Armonies & Hellwig, 1987, and *M. extraculum* Ax & Armonies, 1990. Of these species, however, only *M. leptos*, *M. reynoldsi*, and *M. extraculum* have the characteristic distal enlargement of the stylet which assigns them to the *Macrostromum tuba* group. The other species listed above belong to the *Macrostromum orthostylum* group.

Macrostromum coomerensis, *M. reynoldsi*, *M. extraculum*, and *M. leptos* are differentiated by the different length and outline of their stylets. The stylet of *M. coomerensis* (125 μm in length) is essentially longer than that of both *M. reynoldsi* (72 μm) and *M. extraculum* (68-72 μm). For *M. leptos*, only the length of the body (0.8mm) and the outline of the stylet are known. The stylet of *M. leptos* differs greatly from that of *M. coomerensis* both in the nature of the distal swelling with subterminal pore and in having the proximal and median part of the stylet greatly

curved. Distally, the stylet is only weakly corkscrew-like.

Macrostomum greenwoodi belongs to the *Macrostomum orthostylum* group based on the curved and evenly tapered distal region of the stylet. The main distinguishing character for this species concerns the outline of the distal termination of the stylet. The distal tip in *M. greenwoodi* is obtuse, with a subterminal ejaculatory duct opening. In this respect, and in not having a cork-screw like stylet, the following 9 species resemble with *M. greenwoodi*: *M. curvituba* Luther, 1947, *M. infundibuliferum* Plotnikov, 1905, *M. johni* Young, 1972, *M. longituba* Papi, 1953, *M. lutheri* Beklemishev, 1927, *M. mediterraneum* Ax, 1956, *M. magnacurvituba* Ax, 1994, *M. mosquense* Beklemishev, 1951, and *M. tennicauda* Luther, 1947. With the exception of *M. lutheri* and *M. greenwoodi* all other species have stylets with the ejaculatory duct opening subterminally on the concave side of the stylet. The stylets of *M. greenwoodi* and *M. lutheri* have ejaculatory openings subterminally on the convex side of the stylet. *M. lutheri* was established by Beklemishev (1927) only on the characteristic termination of the stylet. Other morphological characters are unknown. Therefore, the only main differential character resides in the complicate enlargement of the distal termination of the stylet.

ACKNOWLEDGEMENTS

Financial support was given by the Deutsche Forschungsgemeinschaft to Dr A. Faubel. We thank Prof. G. Grigg, Head of Zoology

Department, Prof. Jack Greenwood and staff for facilities and help, University of Queensland, Brisbane. Dr L.R.G. Cannon, Queensland Museum, Brisbane and Prof. K. Rohde, University of New England, Armidale, NSW, assisted with help and arrangements.

LITERATURE CITED

- BEKLEMISHEV, V.N. 1927. Über die Fauna des Aralsees. Zugleich ein Beitrag zur Morphologie und zur Systematik der Dalyellioidea. Zoologisches Jahrbuch, Systematik 54: 87-138.
- BEKLEMISHEV, V.N. 1951. The species of the genus *Macrostomum* (Turbellaria, Rhabdocoela) of the USSR [in Russian]. Bulletin of the natural Society of Moscow (Biology, new series) 56, 31-40.
- DÖRJES, J. 1968. Die Acoela (Turbellaria) der deutschen Nordseeküste und ein neues System der Ordnung. Zeitschrift für zoologische Systematik und Evolutionsforschung 6: 56-452.
- FAUBEL, A., BLOME, D. & CANNON, L.R.G. 1994. Sandy beach meiofauna of eastern Australia (southern Queensland and New South Wales). I. Introduction and Macrostomida (Platyhelminthes). Invertebrate Taxonomy 8: 989-1007.
- HASWELL, W.A. 1905. Studies on Turbellaria. Quarterly Journal of microscopical Science 49: 425-467.
- TRENCH, R.K. & WINSOR, H. 1987. Symbiosis with dinoflagellates in two pelagic flatworms, *Amphiscolops* sp. and *Haplodiscus* sp. Symbiosis 3: 1-22.
- WINSOR, L. 1988. A new acoel (Convolutidae) from the north Queensland coast, Australia. Fortschritte der Zoologie/Progress in Zoology 36: 391-394.
1990. Marine Turbellaria (Acoela) from north Queensland. Memoirs of the Queensland Museum 28: 785-800.